

Regulation of Jak2 Function by Phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ during Cytokine Signaling[∇]

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Jak2, the cognate tyrosine kinase for numerous cytokine receptors, undergoes multisite phosphorylation during cytokine stimulation. To understand the role of phosphorylation in Jak2 regulation, we used mass spectrometry to identify numerous Jak2 phosphorylation sites and characterize their significance for Jak2 function. Two sites outside of the tyrosine kinase domain, Tyr₃₁₇ in the FERM domain and Tyr₆₃₇ in the JH2 domain, exhibited strong regulation of Jak2 activity. Mutation of Tyr₃₁₇ promotes increased Jak2 activity, and the phosphorylation of Tyr₃₁₇ during cytokine signaling requires prior activation loop phosphorylation, which is consistent with a role for Tyr₃₁₇ in the feedback inhibition of Jak2 kinase activity after receptor stimulation. Comparison to several previously identified regulatory phosphorylation sites on Jak2 revealed a dominant role for Tyr₃₁₇ in the attenuation of Jak2 signaling. In contrast, mutation of Tyr₆₃₇ decreased Jak2 signaling and activity and partially suppressed the activating JH2 V617F mutation, suggesting a role for Tyr₆₃₇ phosphorylation in the release of JH2 domain-mediated suppression of Jak2 kinase activity during cytokine stimulation. The phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ act in concert with other regulatory events to maintain appropriate control of Jak2 activity and cytokine signaling.

Type I cytokines act via cell surface receptors on target cells to mediate a plethora of physiologic processes, ranging from hematopoietic and immune functions (such as those controlled by erythropoietin and the interleukins), to growth and neuroendocrine responses (such as those modulated by growth hormone and leptin) (16, 19, 20, 29, 34). Cytokine receptors contain an extracellular ligand-binding domain, a single transmembrane domain, and an intracellular domain that, although devoid of enzymatic activity, transmits intracellular signals by means of an associated Jak family tyrosine kinase. Ligand binding activates the receptor-associated intracellular Jak kinase, resulting in Jak kinase autophosphorylation and activation, as well as the subsequent tyrosine phosphorylation of the intracellular domain of the cytokine receptor. These tyrosine phosphorylation events mediate downstream signaling by the cytokine receptor/Jak kinase complex (16, 20, 23, 29).

The Jak kinase family contains four members: Jak1 to Jak3 and Tyk2 (16, 20). Of these, Jak1-2 and Tyk2 are ubiquitously expressed, while Jak3 is found predominantly in immune and hematopoietic tissues. Jak kinases contain four conserved domains: the NH₂-terminal FERM domain mediates cytokine receptor interactions (36, 39), while function of the adjacent (nonphosphotyrosine binding) SH2-like fold remains unclear. The COOH-terminal region of Jak kinases contains a kinase-like JH2 domain that is devoid of enzymatic activity but which

regulates the activity of the COOH-terminal JH1 tyrosine kinase domain (11, 24, 32, 37, 38).

Our laboratory studies signaling by LepRb, which regulates energy balance, neuroendocrine homeostasis, and immune function in response to leptin, a hormonal signal of long-term energy stores (10, 12, 29, 34). Leptin binding to LepRb promotes the activation and tyrosine phosphorylation of the LepRb-associated Jak2, resulting in the phosphorylation of tyrosine residues on Jak2 and the intracellular tail of LepRb Jak2 (2, 22, 29). Tyrosine phosphorylation sites on LepRb recruit signal transducers and activators of transcription (STATs) and SHP-2 to mediate downstream signaling, as well as the suppressor of cytokine signaling-3 (SOCS3) to attenuate LepRb signaling (2, 5, 29).

Several sites of Jak2 tyrosine phosphorylation have also been identified, and functions for some of these sites have been elucidated: within the FERM domain, phosphorylation of Tyr₁₁₉ disrupts Jak2-cytokine receptor interactions (13). Within and adjacent to the JH2 domain, the phosphorylation of Ser₅₂₃ and Tyr₅₇₀ inhibits Jak2 kinase activity (1, 8). Within the kinase domain itself, phosphorylated Tyr₈₁₃ mediates binding of SH2-B/SH2B1 to increase Jak2 signaling (23), phosphorylation of the activation loop residues Tyr₁₀₀₇ and Tyr₁₀₀₈ plays an essential role in kinase activation (9), and the phosphorylation of Tyr₉₁₃ inhibits Jak2 signaling (15). Other sites of Jak2 phosphorylation also exist (some known, while others have remained undefined), although the function(s) for many of these remain unknown (1, 25). We report here the MS analysis of Jak2 protein, which revealed several novel sites of phosphorylation. We also report the in-depth analysis of two cytokine-regulated Jak2 phosphorylation sites outside of the kinase domain: Tyr₃₁₇ and Tyr₆₃₇. Phosphorylation of Tyr₃₁₇ mediates

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negative-feedback regulation for Jak2, while phosphorylation of Tyr₆₃₇ is necessary for maximal Jak2 kinase activity. We propose an integrated model for how these and other phosphorylation sites orchestrate the activity of Jak2.

MATERIALS AND METHODS

Antibodies, growth factors, and reagents. Antibodies recognizing phosphorylated Tyr₃₁₇ and phosphorylated Tyr₆₃₇ were generated by raising rabbit polyclonal antibodies against these sites. Synthetic peptides corresponding to phosphorylated Tyr₃₁₇ (CODVQLY*CDFPD) and Tyr₆₃₇ (CGSLDTY*LKKNK) motifs were generated in the MDRTC Peptide Core (University of Michigan), conjugated to keyhole limpet hemocyanin, and inoculated into rabbits. Antisera were affinity purified by using a 1:1 mixture of Affigel-10 and Affigel-15 coupled to phosphorylated synthetic peptide. The resulting antibody was further purified by incubating it with Affigel coupled to the nonphosphorylated variant of the peptide in combination with an irrelevant phosphorylated peptide. α Jak2(476) antisera for immunoprecipitation were generated by injection into rabbits of keyhole limpet hemocyanin-coupled synthetic peptide with the sequence DSQR KLFYEDKHLQLPAPK. Generation of α Jak2(1139) for immunoblotting has been described previously [previously referred to as α Jak2(NT)] (21). Antibodies to phosphorylated (activated) extracellular signal-regulated kinase (ERK) and phosphorylated (activated) STAT3 were purchased from Cell Signaling Technology (Danvers, MA). Antibody directed against the phosphorylated activation loop of Jak2 (phospho-Tyr_{1007/8}) was purchased from Upstate Biotechnology (Lake Placid, NY). Antibody against the hemagglutinin (HA) epitope was purchased from Covance (Denver, PA). Antibody to the FLAG epitope was purchased from Sigma. Synthetic erythropoietin (EpoGen) was obtained from Amgen (Thousand Oaks, CA).

Generation of Jak2 constructs. The generation of ELR has been described previously (2). The generation of Jak2/pCDNA3 has been described previously (22). To make epitope-tagged Jak2, DNA encoding either a double HA epitope or a double FLAG epitope was added to the 3' end of the Jak2 open reading frame by using a two-step strategy. In the first stage, PCR was conducted using an oligonucleotide that primed upstream of Jak2's internal XhoI site in conjunction with a long downstream oligonucleotide that contained the epitope tag, an NheI site, and a region homologous to the 3' end of Jak2. The PCR product was inserted into TOPO (Invitrogen). An NheI/XhoI digest was used to move the epitope tag containing fragment back into the original Jak2 plasmid. QuikChange mutagenesis (Stratagene) was used to generate Jak2^{Y317F}, Jak2^{Y637E}, Jak2^{Y317E}, Jak2^{Y637E}, Jak2^{K882E}, Jak2^{Y1007/8F}, Jak2^{Y119E-HA}, Jak2^{Y119E/Y317F-HA}, Jak2^{Y119E/Y637F-HA}, Jak2^{Y317F/S523A}, Jak2^{Y317F/Y570F}, Jak2^{S523A/Y570F}, and Jak2^{Y317F/S523A/Y570F}. The presence of the desired mutations and the absence of adventitious mutations were confirmed by DNA sequencing for all plasmids generated.

Cell lines and transfection. HEK293 cells were grown at 37°C in humidified air with 5% CO₂. Cells were grown in Dulbecco modified Eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS), penicillin, and streptomycin. HEK293 cells were transfected by using Lipofectamine (Invitrogen). At 3 to 5 h after transfection, the cells were serum starved overnight in DMEM supplemented with 0.5% bovine serum albumin. 32D cells were grown in RPMI 1640 medium supplemented with 10% FBS and 5% interleukin-3 (IL-3) conditioned medium. IL-3 conditioned medium was generated by using WEHI cells grown in RPMI 1640 medium supplemented with 10% FBS. Cells were grown to confluence, and the medium was harvested and then filtered (0.2- μ m pore size) and stored at 4°C.

Cytokine stimulation, lysis, and immunoprecipitation. HEK293 cells were stimulated with EpoGen at 12.5 U/ml. 32D cells were stimulated with undiluted IL-3 conditioned medium from WEHI cells. Cells were harvested with lysis buffer (20 mM Tris [pH 7.4], 150 mM NaCl, 1% Nonidet P-40, 2 mM phenylmethylsulfonyl fluoride, 2 mM sodium orthovanadate) and cleared by centrifugation. Lysates used for direct immunoblotting were mixed 1:1 with Laemmli buffer. Total Jak2 was immunoprecipitated using a polyclonal antibody to Jak2 and protein A-agarose (Invitrogen) overnight at 4°C. Immunoprecipitates were washed three times with lysis buffer before elution with Laemmli buffer. HA epitope-tagged Jak2 was immunoprecipitated with a monoclonal antibody to HA and protein G-Sepharose (GE Healthcare) overnight at 4°C. Immunoprecipitates were washed three times with lysis buffer before elution with Laemmli buffer.

MS. For preparation of protein for MS analysis, material was immunoprecipitated from 5 to 10 15-cm dishes of HEK293 cells and resolved by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). Jak2 protein was visu-

alized by Coomassie brilliant blue G-250 stain. Jak2 protein was subject the mass spectrometry (MS) by using two different instruments and methods. In one method, gel slices containing Jak2 were digested with 5 ng of sequencing-grade modified trypsin (Promega)/ μ l in 25 mM ammonium bicarbonate containing 0.01% *n*-octylglucoside for 18 h at 37°C. Peptides were eluted from the gel slices with 80% acetonitrile–1% formic acid. Tryptic digests were separated by capillary high-pressure liquid chromatography (HPLC; C₁₈; 75 μ m [inner diameter] Picofrit column; New Objective) using a flow rate of 100 nl/min over a 3-h reversed-phase gradient and analyzed by using an LTQ two-dimensional linear ion trap mass spectrometer (ThermoFinnigan). Resultant tandem MS (MS/MS) spectra were matched against mouse Jak2 sequence by using TurboSequest (BioWorks 3.1) with a fragment ion tolerance of <0.5 and amino acid modification variables, including phosphorylation (80 Da) of Ser, Thr, and Tyr; oxidation (16 Da) of Met; and methylation (14 Da) of Lys.

For the second method, gel slices containing Jak2 were destained, reduced, and alkylated as described elsewhere (33). Proteolytic digestion was carried out with 13 ng of sequencing-grade trypsin (Promega)/ μ l in 25 mM ammonium bicarbonate for 18 h at 37°C. The resulting peptides were eluted with solution containing 60% methanol–5% acetic acid, and the peptide mixture was lyophilized (Labconco, Kansas City, MO). Esterification and immobilized metal affinity chromatography enrichment were performed with "optimized" buffers described previously (30). Phosphopeptide-enriched peptide mixture was loaded off-line onto precolumn packed with an 8-cm bed length of 5- to 15- μ m spherical C₁₈ beads (Applied Biosystems, Foster City, CA) in 75- μ m (inner-diameter) fused silica capillary tubing. HPLC was performed with a high-performance analytical column (50 μ m by 8 cm) packed with 5- μ m-diameter C₁₈ reversed-phase beads (YMC, Wilmington, NC) (30) at an estimated eluent flow rate of 20 to 50 nl/min. HPLC solvent A was 0.2 M acetic acid, and solvent B was 70% acetonitrile–0.2 M acetic acid. LC-MS analysis of phosphopeptide mixture was performed by using a solvent gradient of 0 to 5% solvent B for 5 min and 5 to 100% solvent B for 35 min. MS data acquisition was performed on QSTAR XL in data-dependent mode (MS scan, 300 $\leq m/z \leq$ 2,000, top five most abundant MS/MS scans using low resolution for precursor isolation and using 1.5-s accumulation with enhance-all mode, 1.8-kV electron spray ionization voltage). The resulting spectra were compared against the NCBI mouse database using the search engine Mascot Daemon (Matrix Science, Inc., London, United Kingdom). The search parameters allowed for two missed cleavages for trypsin; a fixed modification of +14 for *o*-methyl esters, for aspartic acid, glutamic acid, and peptide C terminus; and variable modifications of +80 for serine, threonine, and tyrosine phosphorylation and +16 for methionine oxidation. The mass tolerance was 1.0 Da for precursors and 0.35 Da for fragment ions. MS/MS spectra corresponding to phosphorylated peptides of Jak2 were manually verified.

Immunoblotting. SDS-PAGE gels were transferred to nitrocellulose membranes (Whatman) in Towbin buffer containing 0.02% SDS and 20% methanol. Membranes were blocked for 1 h at room temperature or overnight at 4°C in buffer containing 20 mM Tris (pH 7.4), 150 mM NaCl, and 0.01% Tween 20 (wash buffer) supplemented with 3% bovine serum albumin (block buffer). Membranes were incubated in primary antibody in block buffer for 2 h at room temperature or overnight at 4°C. Membranes were rinsed three times with wash buffer and then incubated with a secondary antibody conjugated to either horseradish peroxidase (Santa Cruz) or a fluorescent tag (Molecular Probes or LI-COR). For horseradish peroxidase detection, membranes were rinsed three times in wash buffer before being treated with chemiluminescence reagent (Roche) and exposed to film (Kodak, Denville, NJ). For fluorescence detection, membranes were rinsed three times in wash buffer before detection using an Odyssey imaging system manufactured by LI-COR Biosciences (Lincoln, NE). Immunoreactive bands were identified and quantified using Odyssey application software version 2.1.

In vitro kinase assays. HEK293 cells were transfected with plasmids encoding ELR and either mutant or wild-type (WT) Jak2. After transfection, the cells were serum starved overnight in DMEM supplemented with 0.5% bovine serum albumin. Prior to lysis, cells were treated with either vehicle or erythropoietin for 15 min. Cells were harvested with lysis buffer (50 mM Tris [pH 7.4], 150 mM NaCl, 2 mM EGTA, 0.2% Triton X-100, 0.5 mM dithiothreitol, 1 mM phenylmethylsulfonyl fluoride, 1 mM Na₃VO₄, 10 μ g of aprotinin/ml, 10 μ g of leupeptin/ml) and cleared by centrifugation. Lysates were immunoprecipitated with an antibody to Jak2 or an antibody to the HA epitope. Immunoprecipitates were washed twice with lysis buffer and then twice with kinase buffer (50 mM HEPES [pH 7.5], 100 mM NaCl, 1 mM Na₃VO₄, 5 mM MnCl₂, 0.5 mM dithiothreitol). Washed immunoprecipitates were split into two fractions. One fraction was used in the kinase assay. The kinase reaction was initiated by adding kinase buffer containing 500 mM STAT5 target peptide (AKAADGYVKPQIKQVV), 125 μ M ATP, and 10 μ Ci of ³²P-labeled γ -ATP to each immunoprecipitate. Aliquots

TABLE 1. Jak2 phosphorylation sites identified by MS/MS in the present study

Residue(s)	Jak2 phosphorylation site ^a	Previous identification ^b	Function ^c
Tyr	134	Novel	NS
	201	18	SHP-2 binding?
	206	18	NS
	221	1, 8, 25	Weakly activating?
	317	Novel	NS
	372	Novel	NS
	423	Psite	NS
	435	Psite	NS
	570	1, 8, 25	Inhibitory
	637	25	NS
	813	23, 25	Activating/SH2B binding
	868	25*	NS
	966	25*	NS
1007/8	9	Activating	
Ser/Thr	523	21, 26	Inhibitory
	668	Novel	NS
	904	Novel	NS

^a That is, the sites identified with high confidence (each site has been observed in multiple runs, conforms to strict confidence standards, and was manually confirmed).

^b References for any previous description of the site are listed, or its novelty is indicated. PSite, reported but not otherwise studied at www.phosphosite.org. *, the subject of a separate study (L. S. Argetsinger and C. Carter-Su, unpublished data).

^c Any known or presumptive function for the site is indicated. NS, not previously studied.

were taken from the kinase reaction at various time points and blotted on P81 papers. The P81 paper was washed in 75 mM H₃PO₄, scintillation fluid (Cytosint [BP Biomedical]) was added, and bound ³²P was counted by using a Packard scintillation counter. The second immunoprecipitate fraction was used to measure Jak2 content used to normalize kinase assay data. This fraction was resolved by SDS-PAGE, transferred to nitrocellulose, and then immunoblotted with a monoclonal antibody directed against the total Jak2 or a monoclonal antibody directed against the HA epitope. Fluorescent secondary antibody was used, and immunoreactive bands were imaged and quantified by using an Odyssey infrared imaging system (LI-COR Biosciences).

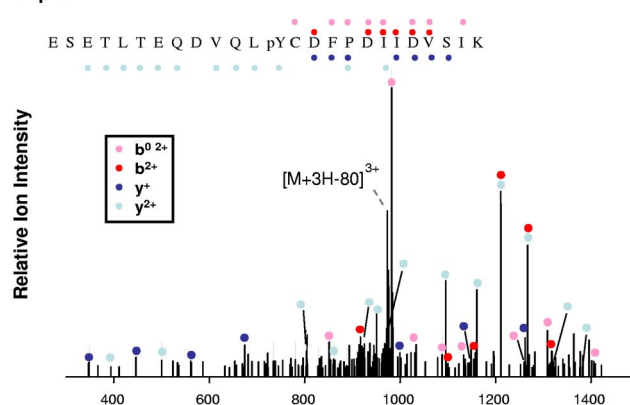
RESULTS

Identification of Jak2 phosphorylation sites. We used MS analysis of Jak2 protein to identify phosphorylation sites on Jak2. We prepared Jak2 by overexpression in HEK293 cells. To enable cytokine-mediated activation, we cotransfected into HEK293 cells plasmids encoding Jak2 and ELR (a chimeric receptor containing the extracellular domain of the erythropoietin receptor and the intracellular domain of LepRb, which effectively places the LepRb intracellular domain under the control of erythropoietin). ELR was utilized because it is expressed more highly and allows more robust activation of Jak2 compared to native LepRb. HEK293 cells expressing Jak2 protein were stimulated with erythropoietin (Epo) for 15 min before lysis and immunoprecipitation to isolate Jak2. Immunoprecipitates were resolved by SDS-PAGE and stained with Coomassie blue to identify Jak2 protein. Jak2 bands were subjected to proteolytic cleavage and analyzed by two MS instruments, a linear ion trap mass spectrometer and a hybrid quadrupole time-of-flight mass spectrometer. Analysis of the MS/MS spectra revealed numerous precursor ions that represented phosphorylated Jak2 peptides, each of which was ob-

served in multiple runs and conformed to strict standards of confidence by software analysis. Some of these had previously been noted in the literature or on scientific websites (1, 8, 9, 18, 21, 23, 25, 26; www.phosphosite.org), while others (three Tyr and two Ser/Thr residues) were novel; most (11 sites overall) had not been functionally characterized (Table 1).

With respect to Tyr₃₁₇ and Tyr₆₃₇, which represent the focus of this report, analysis of MS/MS spectra from the linear trap instrument using TurboSequest revealed a 3+ charged species corresponding to a Tyr₃₁₇-containing Jak2 tryptic peptide ES ETLTEQDVQL(P)YCDFPDIIDVSIK with Xcorr 3.74. The spectra showed an ion consistent with the neutral loss of 80 Da resulting from the loss of HPO₃ from a triply charged phosphotyrosine-containing peptide (Fig. 1A). Analysis of quadrupole time-of-flight MS/MS spectra using Mascot revealed a 2+ charged precursor corresponding to the Tyr₆₃₇-containing Jak2 tryptic peptide FGSLDT(P)YLK with a score of 51. The spectrum also shows an ion consistent with neutral loss of 80 Da resulting from the loss of HPO₃ from the doubly charged

A. pY317



B. pY637

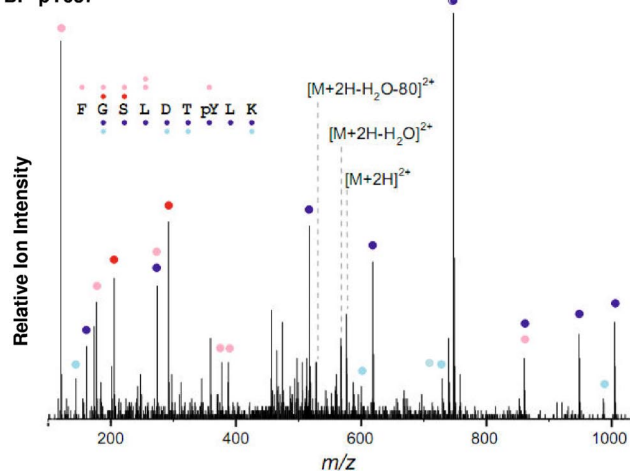


FIG. 1. Representative MS/MS spectra (for pY317 and pY637). MS/MS spectra from Jak2 peptides containing pY317 (A) and pY637 (B) are shown. Sequence assignments of y+ and y+ ions are shown in blue and light blue, respectively. The assignments of b+ ions, and derivative b0+ and a+ ions are presented in red and pink, respectively. An ion corresponding to neutral loss of 80 Da from each precursor is shown.

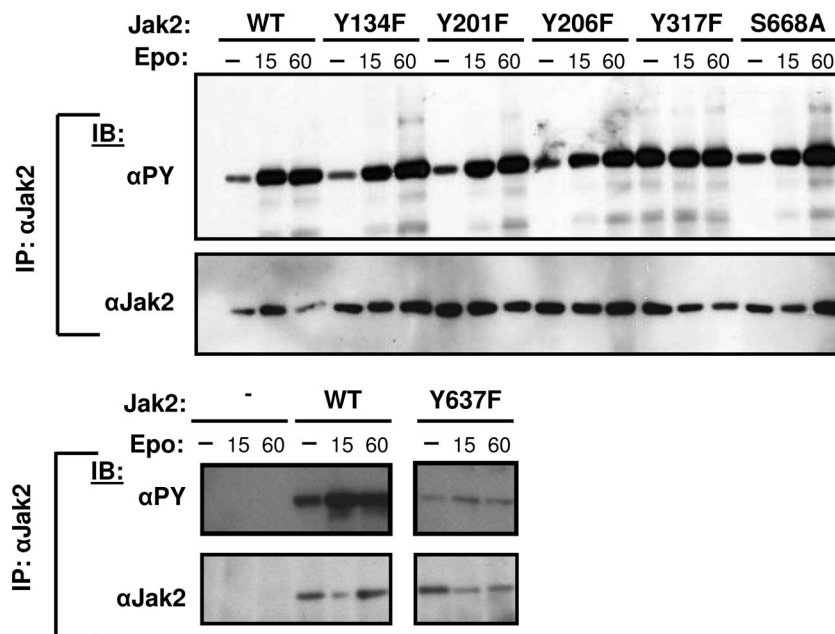


FIG. 2. Representative examples of mutant screening data. HEK293 cells were transfected with the indicated Jak2 or Jak2 mutant plasmid. Cells were made quiescent overnight. Cells were either lysed without treatment (0) or incubated in the presence of Epo (12.5 U/ml) for either 15 or 60 min and then lysed. Lysates were immunoprecipitated (IP) with an antibody to the total Jak2. Immunoprecipitates were resolved by SDS-PAGE, transferred to nitrocellulose, and immunoblotted (IB) with the indicated antibodies. The results shown are typical of three independent experiments.

phosphotyrosine-containing peptide (Fig. 1B). Manual inspection and interpretation of fragmentation spectra confirmed peptide sequences in these and the other noted cases.

Screening candidate phosphorylation sites for functional relevance. In order to determine which of the identified Jak2 phosphorylation sites merited in-depth study, we mutated each site to a nonphosphorylatable residue (using either a Tyr→Phe or Ser→Ala substitution) to create a panel of phosphorylation site-defective Jak2 molecules and examined the basal and hormone-stimulated tyrosine phosphorylation of each Jak2 mutant after coexpression with ELR in HEK293 cells (Fig. 2). While antiphosphotyrosine (α PY) immunoreactivity in immunoblotting represents an indirect measure of Jak2 function, multiple studies have revealed a correlation with Jak2 activity (7, 9), and this analysis should reveal any alterations in Jak2 activity resulting from the loss of a specific phosphorylation site. This analysis revealed two mutants for phosphorylation sites lying outside of the kinase domain that differed significantly from WT Jak2 with respect to α PY immunoreactivity. Jak2^{Y317F} exhibited high α PY immunoreactivity in both stimulated and unstimulated states, suggesting that phosphorylation at Tyr₃₁₇ (a novel site of Jak2 phosphorylation) inhibits Jak2 activity (Fig. 2). Jak2^{Y637F}, conversely, exhibited reduced α PY immunoreactivity in both stimulated and unstimulated states, suggesting that phosphorylation at Tyr₆₃₇ (a previously identified phosphorylation site that has not been functionally studied [25]) is required for maximal Jak2 activity (Fig. 2). Based on these screening data, Tyr₃₁₇ and Tyr₆₃₇ were chosen for further study. A number of sites within the kinase domain also modulate Jak2 tyrosine phosphorylation; the in-depth analysis of these sites will be reported elsewhere (C. Carter-Su and L. S. Argetsinger, unpublished data).

Phosphorylation of Jak2 Tyr₃₁₇ and Tyr₆₃₇ during cytokine stimulation. To examine the phosphorylation status of these sites under different conditions, we generated antibodies against phosphorylated Tyr₃₁₇ and phosphorylated Tyr₆₃₇ (α pY317 and α pY637, respectively). To examine the phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ and the function of the Jak2 molecules mutated at these sites more thoroughly, we coexpressed ELR with either Jak2, Jak2^{Y317F} or Jak2^{Y637F} in HEK293 cells and stimulated them with Epo for various times before lysis. Cell lysates or Jak2 immunoprecipitates were immunoblotted with α PY, α pY1007/8 (which recognizes the phosphorylated activation loop of Jak2), α Jak2, α PY, and α pY317 or α pY637 (Fig. 3).

With respect to Jak2^{Y317F}, we found high baseline α PY and α pY1007/8 immunoreactivity, which increased only slightly upon stimulation and remained elevated for the duration of the experiment (Fig. 3A). This contrasts with Jak2, in which α PY and α pY1007/8 immunoreactivity increased quickly and peaked in the first 30 min of stimulation before slowly decreasing over the course of several hours. The minimal regulation of the tyrosine phosphorylation of Jak2^{Y317F} by Epo may suggest that this Jak2 mutant is close to maximal tyrosine phosphorylation at baseline, limiting further increases with cytokine stimulation. The failure of α pY317 to react with Jak2^{Y317F} suggests the specificity of this antibody preparation for phosphorylated Tyr₃₁₇. Phosphorylation at Tyr₃₁₇ on Jak2, detected by α pY317 immunoblotting, peaked later than tyrosine phosphorylation or activation loop phosphorylation and remained elevated for at least 8 h after stimulation, suggesting a potential role for the phosphorylation of Tyr₃₁₇ in the feedback inhibition of Jak2 activity following cytokine stimulation.

With respect to Jak2^{Y637F}, overall α PY and α pY1007/8 im-

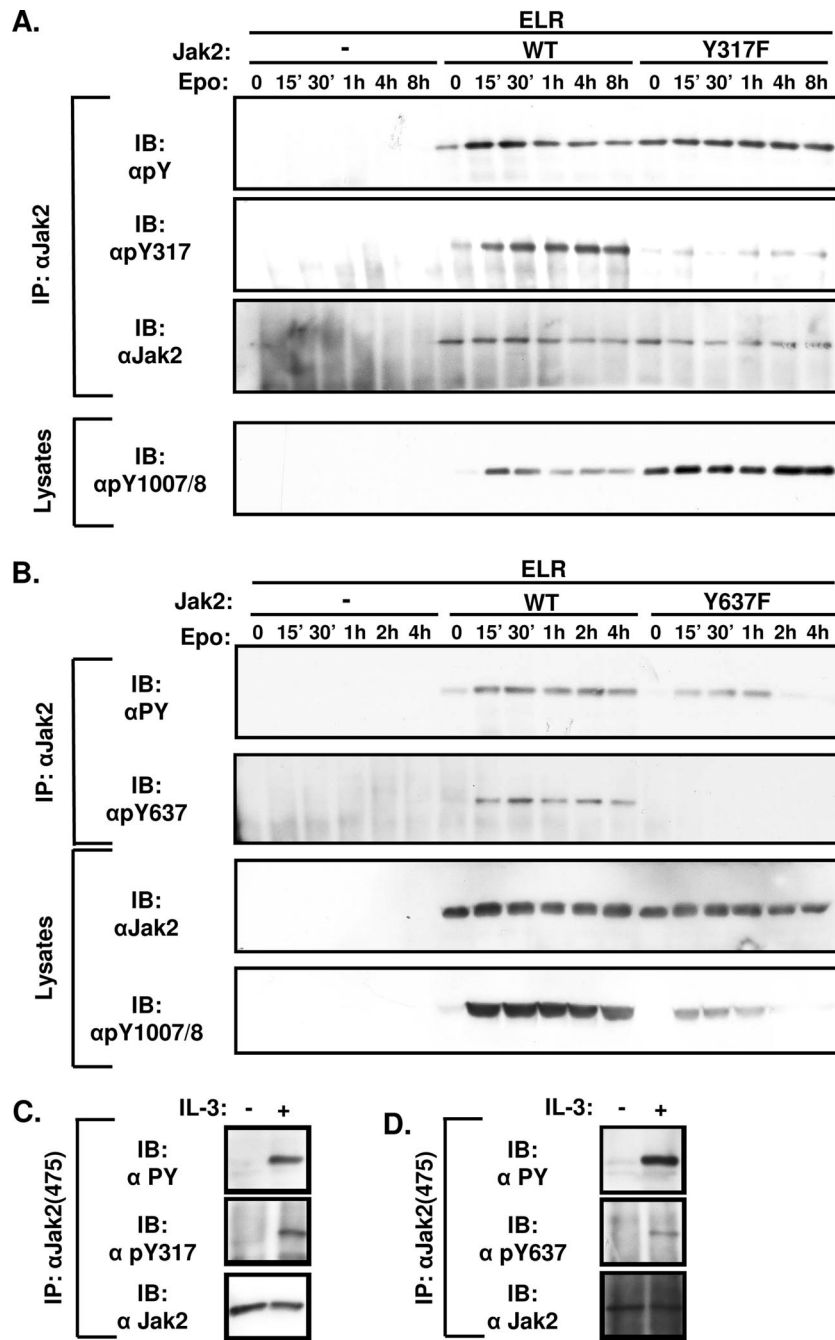


FIG. 3. Time course of phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ after cytokine stimulation. (A and B) HEK293 cells were transfected with ELR and the indicated Jak2 isoform, made quiescent, and incubated with Epo (12.5 U/ml) for the indicated amounts of time before being lysed. Lysates were immunoprecipitated (IP) with the indicated antibodies. Lysates or immunoprecipitated proteins (as indicated) were resolved by SDS-PAGE and transferred to nitrocellulose for immunoblotting (IB) with the indicated antibodies. The figures shown are typical of multiple independent experiments. (C and D) 32D cells were made quiescent for 4 h and then incubated with either serum-free medium or undiluted IL-3 containing WEHI cell conditioned medium for 1 h (C) or 30 min (D) before lysis. Lysates were immunoprecipitated with an antibody to the total Jak2. Immunoprecipitates were resolved by SDS-PAGE, transferred to nitrocellulose, and blotted with the indicated antibodies. The results shown are representative of multiple independent experiments.

munoreactivity was decreased compared to Jak2 at baseline and over the entire time course of stimulation (Fig. 3B). The stimulation of αPY₆₃₇ reactivity on Jak2 by cytokine treatment and the failure of αPY₆₃₇ to react with Jak2^{Y637F} revealed the specificity of this antibody for phosphorylated Tyr₆₃₇. The

phosphorylation of Tyr₆₃₇ increased rapidly during the first 30 min of stimulation before declining (as for overall αPY and αPY_{1007/8} immunoreactivity), suggesting a potential role for the phosphorylation of Tyr₆₃₇ in the activation of Jak2 during cytokine signaling. We also observed IL-3-stimulated phosphor-

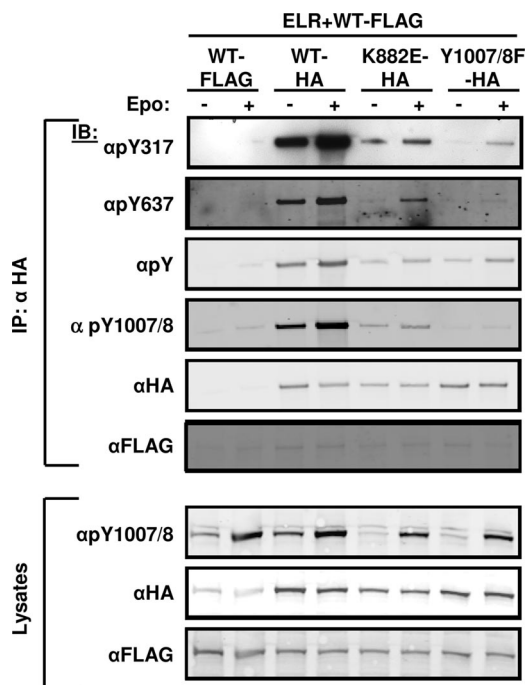


FIG. 4. Transphosphorylation of Tyr₃₁₇ and Tyr₆₃₇ in response to cytokine stimulus. HEK293 cells were transfected with the indicated plasmids, made quiescent overnight, and incubated in the absence (–) or presence (+) of Epo (12.5 U/ml) for 15 min before lysis. Lysates were immunoprecipitated (IP) with α HA antibody. Immunoprecipitates or lysates (as indicated) were resolved by SDS-PAGE, transferred to nitrocellulose, and immunoblotted (IB) with the indicated antibodies. The results shown are typical of multiple independent experiments.

ylation of Tyr₃₁₇ and Tyr₆₃₇ on endogenous Jak2 purified from 32D myeloid progenitor cells (21, 35) (Fig. 3C and D), suggesting the phosphorylation of these sites in response to multiple cytokines.

Transphosphorylation of Tyr₃₁₇ and Tyr₆₃₇ during cytokine stimulation. Protein autophosphorylation is often ordered and can occur by intramolecular (*cis*) or intermolecular (*trans*) mechanisms. To determine the potential mechanisms underlying phosphorylation at Tyr₃₁₇ and Tyr₆₃₇ and their timing relative to the activating Tyr_{1007/1008} phosphorylation, we generated HA epitope-tagged Jak2 (Jak2-HA) and two different inactive variants of Jak2-HA. The mutation in Jak2-HA^{K882E} blocks ATP binding to the kinase domain, while the alteration in Jak2-HA^{Y1007/8F} blocks the phosphorylation of the activation loop in the Jak2 kinase domain. We coexpressed these Jak2-HA constructs with ELR plus a FLAG epitope-tagged WT Jak2 (Jak2-FLAG) and used α HA to immunoprecipitate the Jak2-HA isoforms in order to assess their phosphorylation status with phospho-specific antibodies (Fig. 4). The finding of similarly insignificant amounts of α HA-precipitable α FLAG-reactive Jak2 under each condition (including without Jak2-HA isoforms [see overexposed α FLAG panel in Fig. 4]) revealed that the signal from each phospho-specific antibody is attributable to the precipitated Jak2-HA isoform. Although the phosphorylation of Jak2-HA on Tyr_{1007/8}, Tyr₃₁₇ and Tyr₆₃₇ was consistently greater than for Jak2-HA^{K882E}, each of

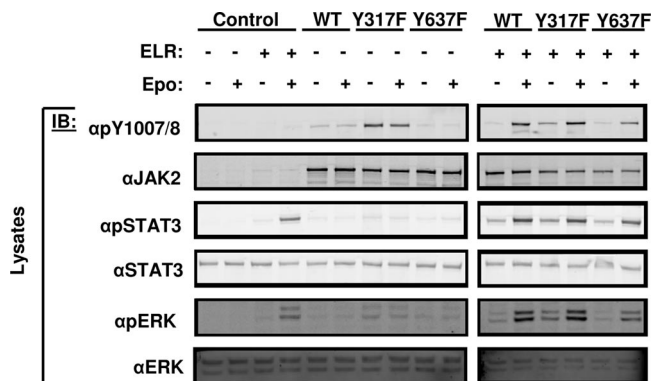


FIG. 5. Effects of Tyr₃₁₇ and Tyr₆₃₇ on downstream signaling. HEK293 cells were transfected with the indicated plasmids, made quiescent overnight, and incubated in the absence (–) or presence (+) of Epo (12.5 U/ml) for 15 min before lysis. Lysates were resolved by SDS-PAGE, transferred to nitrocellulose, and immunoblotted (IB) with the indicated antibodies. The results shown are typical of multiple independent experiments.

these sites was phosphorylated on the ATP-binding mutant, a finding consistent with the transphosphorylation of these sites by Jak2-FLAG (Fig. 4). The decreased phosphorylation of Jak2-HA^{K882E} relative to Jak2-HA could reflect a combination of absent *cis* autophosphorylation of Jak2-HA^{K882E} (which might occur in Jak2-HA), decreased total kinase activity in Jak2-HA^{K882E}-expressing cells, and/or an altered conformation of the inactive Jak2-HA^{K882E}. Interestingly, the decreased phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ on Jak2-HA^{K882E} was more pronounced than the decrease in total α PY reactivity: while α PY reactivity per unit of HA-immunoreactive protein on stimulated Jak2-HA^{K882E} was decreased to ca. 40% of Jak2-HA levels, phosphorylation at Tyr₃₁₇ and Tyr₆₃₇ on stimulated Jak2-HA^{K882E} was decreased to ca. 5 and 10% of Jak2-HA levels, respectively. This difference suggests a stricter requirement for Jak2 kinase activity in order to phosphorylate Tyr₃₁₇ and Tyr₆₃₇ compared to other major sites of tyrosine phosphorylation on Jak2. In addition, in Jak2-HA^{Y1007/8F}, which cannot undergo the conformational change associated with activation loop phosphorylation, the phosphorylation of both Tyr₃₁₇ and Tyr₆₃₇ was further decreased to ca. 1% compared to Jak2-HA (relative to HA-immunoreactive protein and relative to total α PY immunoreactivity). Thus, these data are consistent with the transphosphorylation of Tyr₃₁₇ and Tyr₆₃₇ and also with the requirement for Tyr_{1007/8}- and activity-dependent conformational changes to enable the phosphorylation of Tyr₃₁₇ and Tyr₆₃₇ during Jak2 activation, placing the phosphorylation of these sites downstream of the initial phosphorylation event of kinase activation.

Effects of Tyr₃₁₇ and Tyr₆₃₇ on cytokine signaling. LepRb/Jak2 signaling promotes the activation of STAT3 via the phosphorylation of Tyr₁₁₃₈ on LepRb, and mediates the activation of ERK both via the phosphorylation of Tyr₉₈₅ on LepRb and via Jak2-mediated signals that are independent of LepRb phosphorylation sites (3, 4). To determine the potential roles for Tyr₃₁₇ or Tyr₆₃₇ on these downstream pathways, we measured the activation of ERK and STAT3 by assessing their phosphorylation in cells expressing Jak2, Jak2^{Y317F}, or Jak2^{Y637F} in the presence or absence of ELR (Fig. 5). Consis-

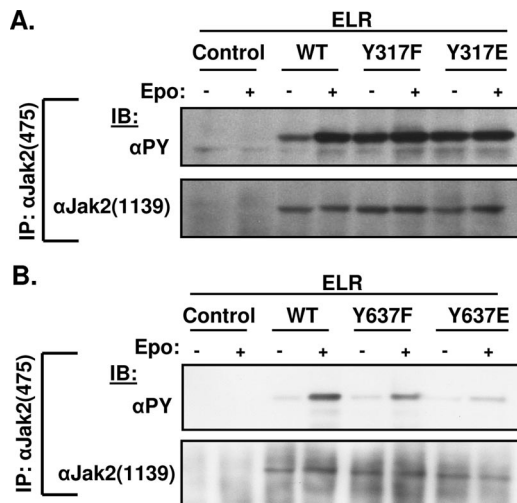


FIG. 6. Similar phenotypes of Phe and Glu substitution mutants for Tyr₃₁₇ and Tyr₆₃₇. (A and B) HEK293 cells were transfected (IP) with the indicated plasmids, made quiescent overnight, and incubated in the presence or absence of Epo (12.5 U/ml) before lysis. Lysates were immunoprecipitated (IP) with the indicated antibody. Immunoprecipitates or lysates (as indicated) were resolved by SDS-PAGE, transferred to nitrocellulose, and immunoblotted (IB) with the indicated antibodies. The figures shown are typical of multiple independent experiments.

tent with its increased α pY1007/8 immunoreactivity, Jak2^{Y317F} mediated increased downstream signaling to STAT3 and ERK compared to Jak2 in the presence of ELR. Across multiple experiments, Jak2^{Y317F} mediated an ~2-fold increase in phosphorylated STAT3 and phosphorylated ERK in unstimulated cells ($n = 3$, $P < 0.05$). The levels of these activated proteins were not significantly different in stimulated cells. In contrast to Jak2^{Y317F}, Jak2^{Y637F} exhibited less α pY1007/8 immunoreactivity and downstream STAT3 and ERK signaling than Jak2. Across multiple experiments, Jak2^{Y637F} mediated an ~40% decrease in phosphorylated STAT3 and phosphorylated ERK in stimulated cells ($n = 3$, $P < 0.05$). The levels of these activated proteins were not significantly different in unstimulated cells. Taken together, these data suggest that alterations in STAT3 and ERK activation are likely the result of changes in Jak2 kinase activity rather than any pathway-specific interactions that the phosphorylation at Tyr₃₁₇ or Tyr₆₃₇ might have on these pathways.

Failure of negatively charged mutations of Tyr₃₁₇ and Tyr₆₃₇ to permit normal Jak2 activity. Tyrosine phosphorylation may regulate protein function in many ways, including by the alteration of protein conformation by the addition of a phosphate group. In order to determine whether this mechanism might operate for Tyr₃₁₇ and/or Tyr₆₃₇, we replaced these residues with the negatively charged amino acid Glu to generate Jak2^{Y317E} and Jak2^{Y637E} and studied their activity relative to Jak2 and their respective Tyr→Phe mutants (Fig. 6). We found that Jak2^{Y317E}, like Jak2^{Y317F}, was highly active in both stimulated and unstimulated states (Fig. 6A). Furthermore, Jak2^{Y637E} was less active than Jak2 and exhibited similar or decreased activity compared to Jak2^{Y637F} (Fig. 6B). Thus, the addition of a negative charge at these sites is not sufficient to mimic the effects

of tyrosine phosphorylation. Neither are the altered activities of Jak2^{Y317F} and Jak2^{Y637F} likely due to nonspecific effects resulting from the substitution of hydrophobic Phe in place of Tyr, since both Glu and Phe produce the same phenotype at each of these sites.

Regulation of Jak2 kinase activity by Tyr₃₁₇ and Tyr₆₃₇. While α PY and α pY1007/8 immunoreactivity generally correlate well with Jak2 kinase activity, this relationship breaks down to some extent at high levels of overall Jak2 phosphorylation, where further increases in phosphorylation become difficult to detect. To gain a more accurate indication of enzymatic activity, we directly measured the *in vitro* kinase activity of immunoaffinity-purified Jak2 and Jak2^{Y317F} toward a target peptide derived from STAT5B, a known substrate of Jak2 (1, 17) (Fig. 7A). These data demonstrated increased *in vitro* kinase activity for purified Jak2^{Y317F} compared to Jak2. Importantly, these data also show that the Jak2^{Y317F} tyrosine kinase, while highly active at baseline, remains responsive to Epo stimulation, suggesting that the mutation of this site does not uncouple kinase function from receptor-mediated regulation. Jak2^{Y637F} displayed reduced kinase activity compared to Jak2 in the stimulated state (Fig. 7B). Jak2^{Y637F} also undergoes activation upon ligand stimulation in the presence of ELR, however. These data confirm the ability of ELR stimulation to increase kinase activity and downstream signaling by Jak2^{Y317F} and Jak2^{Y637F}, suggesting that neither of these sites is required for functional regulation by cytokine receptors.

Regulation of Jak2 by Tyr₃₁₇ and Tyr₆₃₇: a role for cytokine receptor interaction. In addition to the modulation of Jak2 activity by ligand binding to Jak2-associated receptors, the association of Jak2 with cytokine receptors promotes Jak2 activity (baseline Jak2 activity increases in the receptor-bound, relative to the unbound, state), presumably by modulating the conformation of Jak2 (13, 14). The phenotypes of Jak2^{Y317F} and Jak2^{Y637F} are independent of ELR expression in 293 cells (Fig. 5), but it is not clear whether endogenous cytokine receptors in HEK293 cells bind to Jak2 in the absence of ELR to permit the manifestation of potential receptor-dependent effects on signaling.

To determine whether the roles for Tyr₃₁₇ and Tyr₆₃₇ in the regulation of Jak2 function might be independent of cytokine receptor-FERM domain interactions, we examined Jak2^{Y317F} and Jak2^{Y637F} activity in the context of Jak2^{Y119E} (Jak2^{Y119E/Y317F} and Jak2^{Y119E/Y637F}). Phosphorylation of Tyr₁₁₉ disrupts Jak2-FERM domain interaction to most Jak2-binding cytokine receptors, and a Tyr→Glu substitution at Tyr₁₁₉ mimics this effect (13, 14). We thus compared the kinase activity of Jak2^{Y119E}, Jak2^{Y119E/Y317F}, and Jak2^{Y119E/Y637F} to determine whether FERM domain-cytokine receptor interactions were required for modulation of Jak2 activity by Tyr₃₁₇ or Tyr₆₃₇. We found that the activating effect of Tyr₃₁₇ mutation was eliminated in the context of Jak2^{Y119E}, while the inhibitory response to alteration of Tyr₆₃₇ remained in Jak2^{Y119E/Y637F} (Fig. 7C). This suggests that the FERM domain-mediated interaction between Jak2 and cytokine receptors is necessary for the negative regulation of Jak2 by phosphorylation at Tyr₃₁₇ (presumably reflecting a requirement for FERM domain conformation), while the effect of Tyr₆₃₇ on Jak2 kinase activity occurs independently of Tyr₁₁₉. Since Jak2^{Y119E} is still capable of binding gamma interferon receptors (13), it is possible that some

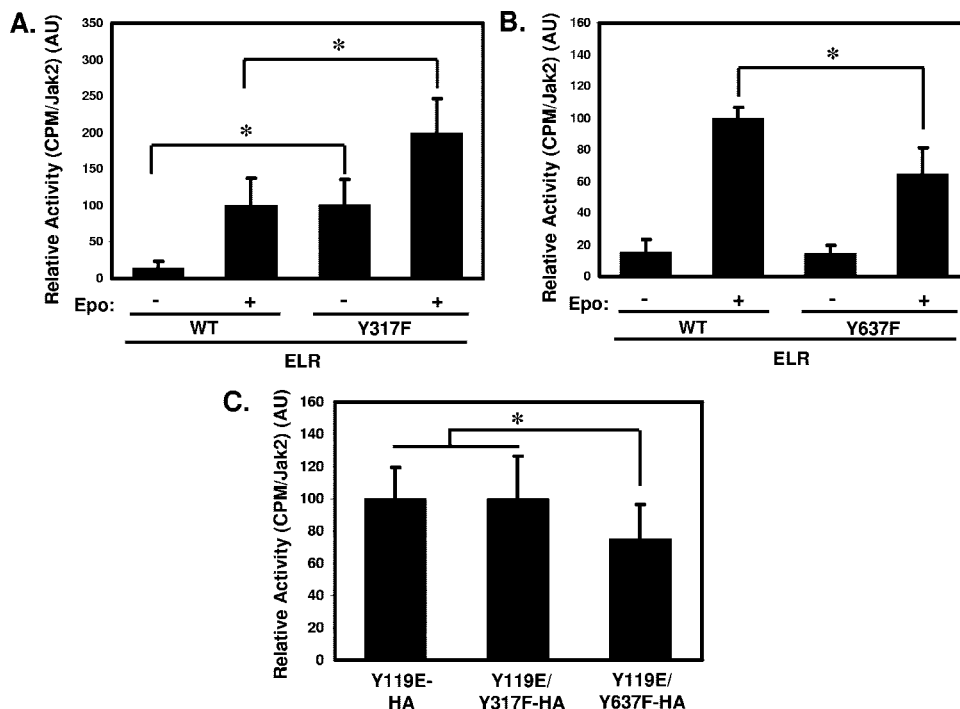


FIG. 7. Modulation of Jak2 kinase activity by Tyr₃₁₇ and Tyr₆₃₇. (A and B) HEK293 cells were transfected with the indicated plasmids made quiescent overnight before treatment with Epo or vehicle and then lysed. Lysates were immunoprecipitated with an antibody to the total Jak2. Immunoprecipitates were subjected to *in vitro* kinase assays using a target peptide derived from STAT5B. Scintillation counts were normalized by Jak2 content (determined by immunoblotting aliquots of each sample in parallel with the kinase assay) and are plotted in arbitrary units (AU). Bars represent the mean value derived from two independent experiments with two replicates each ($n = 4$, total) \pm the standard deviations. Values from each experiment were normalized by the mean value of the Jak2-stimulated samples, which was set at 100 for each experiment. *, $P < 0.05$ (Student unpaired *t* test). (C) HEK293 cells were transfected with the indicated plasmids made quiescent overnight and then lysed. Lysates were immunoprecipitated with α HA. Immunoprecipitates were subjected to *in vitro* kinase assays using a target peptide derived from STAT5B. Scintillation counts were normalized by HA content in each immunoprecipitate and are plotted in AU. Bars represent the mean value derived from three independent experiments with four replicates each ($n = 12$, total) \pm the standard deviations. Values from each experiment were normalized by the mean value of the Jak2^{Y119E} samples, which was set at 100 for each experiment. *, $P < 0.05$ (Student unpaired *t* test).

residual receptor/Jak2^{Y119E} binding could contribute to Tyr₆₃₇ mutation-induced decrements in kinase activity, although the ability of the Tyr₁₁₉→Glu substitution to abrogate the activating effect of Tyr₃₁₇ mutation suggests the likely independence of Tyr₆₃₇ function from FERM domain requirements.

Functional interaction of Tyr₃₁₇ and Tyr₆₃₇ with other regulatory sites on Jak2. Tyr₃₁₇ represents the third inhibitory phosphorylation site that we have identified on Jak2, since phosphorylation at Ser₅₂₃ and Tyr₅₇₀ also inhibits Jak2 (1, 8, 21, 26). To determine the relative contributions of these sites to Jak2 activity, we compared the kinase activity of Jak2^{Y317F}, Jak2^{S523A}, Jak2^{Y570F}, and several Jak2 isoforms containing various combinations of mutations at these inhibitory sites (Fig. 8A to C). The cytokine-stimulated activity of each of these mutants was near maximal and relatively similar (data not shown), and we therefore focused on the kinase activity at baseline in the absence of cytokine stimulation. Jak2^{S523A}, while more active than Jak2 (21, 26), was less active than Jak2^{Y317F}, suggesting a stronger effect of Tyr₃₁₇-mediated inhibition than that of Ser₅₂₃-mediated inhibition (Fig. 8A). Mutation at both sites produced additive effects, however, suggesting the independent effects of these phosphorylation sites on Jak2 activity. The activities of Jak2^{Y317F} and Jak2^{Y570F} were increased relative to Jak2 and similar in magnitude; the addi-

tive effects of mutation at these sites also suggested independent effects on Jak2 activity (Fig. 8B). Comparing the kinase activity of each double mutant with the Jak2^{Y317F/S523A/Y570F} triple mutant revealed that that the most active Jak2 species contain the Tyr₃₁₇ mutation, and the triple mutant demonstrated no increased activity compared to Y317F-containing double mutants, underlining the importance of Tyr₃₁₇ for the appropriate suppression of Jak2 activity (Fig. 8C).

Somatic mutation that substitutes Phe in place Val₆₁₇ of Jak2 (Jak2^{V617F}) mediates the constitutive activation of Jak2 that underlies most cases of polycythemia vera and some portion of other myeloproliferative disorders (28). Val₆₁₇, like Tyr₆₃₇, lies in the JH2 pseudokinase domain of Jak2, and their close proximity suggested the potential for functional interactions between the two residues in the regulation of JH2-mediated control of kinase activity. To determine the ability of Tyr₆₃₇ mutation to suppress the activity of Jak2^{V617F}, we compared the kinase activities of Jak2, Jak2^{V617F}, Jak2^{Y637F}, and Jak2^{V617F/Y637F}. Interestingly, Jak2^{V617F/Y637F} is less active than Jak2^{V617F}, demonstrating that Tyr₆₃₇ influences kinase activity in Jak2^{V617F} (Fig. 8D). Although Jak2^{V617F/Y637F} is less active than Jak2^{Y637F}, it remains more active than Jak2 at baseline, suggesting that Tyr₆₃₇ contributes to only a portion of Val₆₁₇-mediated Jak2 activation.

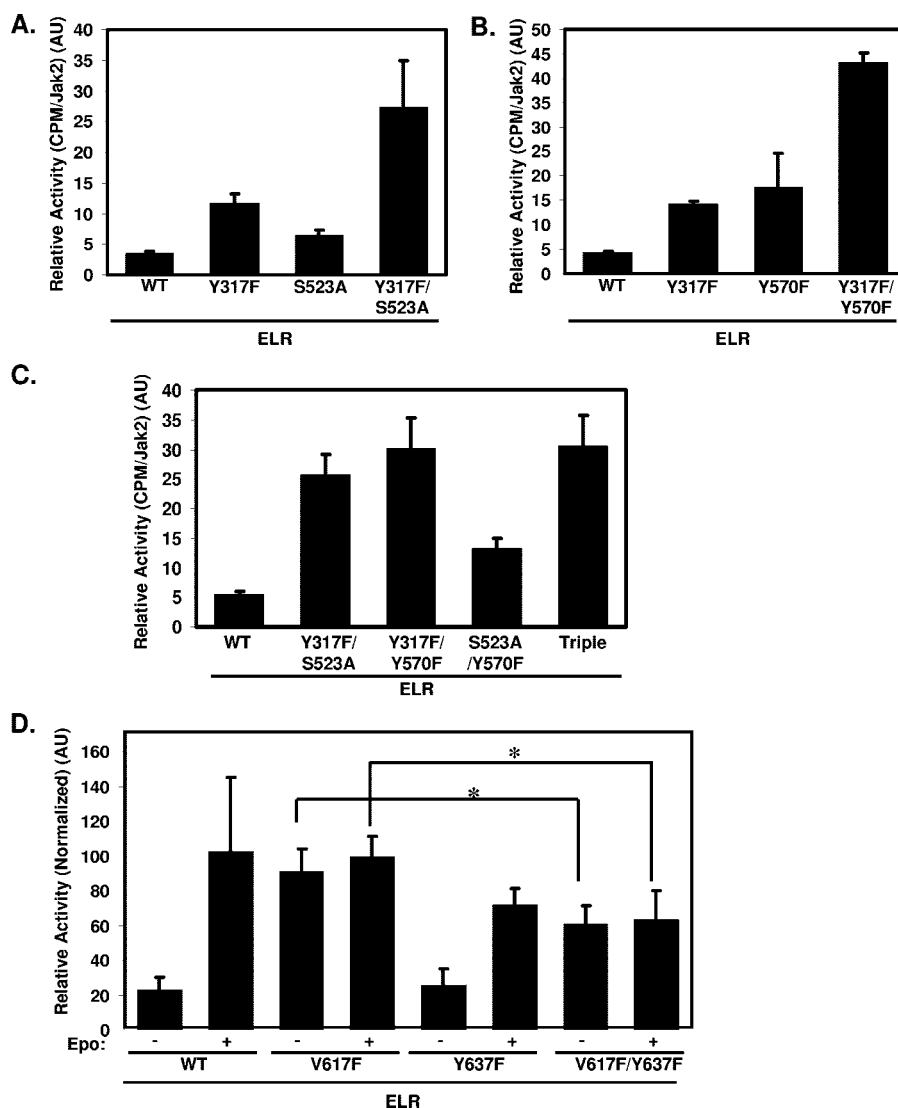


FIG. 8. Functional interaction of Tyr₃₁₇ and Tyr₆₃₇ with other regulatory sites on Jak2. (A to C) HEK293 cells were transfected with the indicated plasmids made quiescent overnight and then lysed. Lysates were immunoprecipitated with an antibody to the total Jak2. Immunoprecipitates were subjected to in vitro kinase assays using a target peptide derived from STAT5B. Scintillation counts were normalized by the Jak2 content in each immunoprecipitate and are plotted in AU. Bars indicate the mean of duplicate samples ($n = 2$). Error bars indicate the standard deviations. (D) HEK293 cells were transfected with the indicated plasmids, made quiescent overnight, and incubated in the presence or absence of Epo (12.5 U/ml) before lysis. Lysates were immunoprecipitated with an antibody to the total Jak2. Immunoprecipitates were subjected to in vitro kinase assays using a target peptide derived from STAT5B. Scintillation counts were normalized by Jak2 content in each immunoprecipitate and are reported in AU. Bars represent the mean values derived from two independent experiments with two replicates each ($n = 4$, total) \pm the standard deviations. Values from each experiment were normalized by the mean value of the Jak2^{V617F}-stimulated samples, which was set at 100. *, $P < 0.05$ (Student unpaired t test).

DISCUSSION

Although many sites of Jak2 phosphorylation have been identified, previous analyses have suggested the presence of others; the function of most Jak2 phosphorylation sites has also remained unclear. Since the analysis of in vitro autophosphorylated Jak2 may not reveal the full complement of physiologically important phosphorylation sites (especially Ser/Thr phosphorylation sites), we utilized MS analysis of Jak2 protein prepared from mammalian cells following cytokine receptor activation in order to identify phosphorylation sites on Jak2. This analysis revealed 18 phosphorylation sites on Jak2, including 15 Tyr phosphorylation sites and 3 Ser/Thr phosphoryla-

tion sites. Of these, 12 sites had been previously published by ourselves or others (1, 8, 9, 18, 21, 23, 25, 26), and two had been presumptively identified by MS analysis reported (www.phosphosite.org). Thus, our present analysis identified three novel sites of Tyr phosphorylation and two novel sites of Ser/Thr phosphorylation on Jak2. The MS data for each of the sites we report here possessed high confidence scores by initial software analysis, and their MS/MS spectra were manually verified and generally showed specific evidence of phosphorylation, such as fragment ions consistent with neutral phosphate loss. Our analysis did not identify some previously reported sites of phosphorylation, including Tyr₁₁₉ in the FERM do-

main and Tyr₉₇₂ (as well as a number of other presumptive sites) in the kinase domain: although spectra potentially consistent with many of these sites were noted, they did not pass our rigorous criteria for inclusion, presumably due to the behavior of the peptides on MS/MS analysis.

We initially assayed the potential importance of these sites by analyzing the basal and ligand-stimulated tyrosine phosphorylation of Jak2 molecules mutant for each identified site and focused our subsequent functional analysis on the two sites outside of the kinase domain that altered Jak2 activity when mutated: Tyr₃₁₇ and Tyr₆₃₇. Additional sites within the kinase domain also regulate Jak2 kinase activity (including Tyr₈₆₈, Tyr₉₆₆, and Tyr₉₇₂); these represent the topic of another study (Carter-Su and Argetsinger, unpublished). Since our screening assay most sensitively detects large changes in Jak2 activity, we cannot rule out more modest roles in Jak2 regulation for additional sites. Since this screen focused on Jak2 phosphorylation, it is also possible that some of the identified sites may play other, unknown, roles in downstream signal transmission for which no assays currently exist.

While cytokine receptor stimulation mediates the phosphorylation of both Tyr₃₁₇ and Tyr₆₃₇, these residues oppositely regulate Jak2-dependent signaling: the mutation of Tyr₃₁₇ enhances Jak2 function, suggesting a role for the phosphorylation of Tyr₃₁₇ in the inhibition of Jak2. Conversely, mutation of Tyr₆₃₇ reduces Jak2 signaling, suggesting a role for the phosphorylation of this residue in the activation of Jak2. Tyr₆₃₇ lies in the highly conserved pseudokinase domain of Jak2, and a corresponding residue is also present in Jak3, suggesting potential partial conservation of the function of this site across the Jak kinase family. In contrast, Tyr₃₁₇ is not conserved with any other member of the Jak family of kinases, suggesting the mechanism by which Tyr₃₁₇ affects protein function is Jak2 specific.

With respect to both Tyr₃₁₇ and Tyr₆₃₇, the functional equivalence of Glu substitution of either of these sites compared to Phe indicates that the negatively charged phosphate in the context of Tyr is required for the function of both Tyr₃₁₇ and Tyr₆₃₇, suggesting that criteria stricter than a simple negative charge must be met to mediate the presumptive intramolecular interactions that underlie the function of these sites. The finding that Glu and Phe substitutions mediate similar effects and the mutants are regulated by cytokine stimulation in a manner proportional to their baseline activity also suggests that non-specific disruption of Jak2 domain structure does not underlie the phenotype of these mutants.

Our data suggest that the interaction of Jak2 with cytokine receptors and/or the conformation of the FERM domain is required for the modulation of Jak2 activity by Tyr₃₁₇, which is consistent with the location of Tyr₃₁₇ in the FERM domain. The decreased activity of free Jak2 (e.g., Jak2^{Y119E}) compared to receptor-associated Jak2 (6, 13, 31), the ability of receptor stimulation to promote the increased activity of Jak2^{Y317F}, and the placement of Tyr₃₁₇ in the extreme COOH terminus of the FERM domain (beyond the presumptive receptor interaction motifs) suggest that Tyr₃₁₇ is unlikely to act by altering the stability of the receptor/Jak2 complex. It is more likely that receptor interaction and/or the conformation of the FERM domain regulates the phosphorylation or function of Tyr₃₁₇; this is consistent with the dependence of Tyr₃₁₇ phosphoryla-

tion on the prior phosphorylation of Tyr_{1007/1008} in the activation loop of the kinase domain, which suggests the requirement for a specific, activated, conformation of Jak2 for the phosphorylation of Tyr₃₁₇. Indeed, we did not detect Tyr₃₁₇ phosphorylation on Jak2^{Y119E} (data not shown), although we cannot rule out the possibility that any Tyr₃₁₇ phosphorylation on this poorly active protein was below our detection threshold.

Our data are thus consistent with a role for Tyr₃₁₇ in the attenuation of Jak2 activity following receptor activation, rather than in suppressing inactive Jak2. Indeed, the phosphorylation of Tyr₃₁₇ is very low in unstimulated Jak2, increases significantly with stimulation, and is highest after full activation of Jak2. Furthermore, the phosphorylation of Tyr_{1007/1008} and the adoption of an activated conformation are required even for the transphosphorylation of Tyr₃₁₇ by an activated Jak2 molecule. Hence, the increased baseline activity of Jak2^{Y317F} in cells prior to cytokine stimulation may result from the inability to inactivate the normally small portion of Jak2 that becomes activated in quiescent HEK293 cells; the failure to return Jak2^{Y317F} to the pool of inactive Jak2 would eventually lead to the accumulation of Jak2 protein that is locked in the active state.

Mutation of Tyr₆₃₇ inhibits kinase activity under most of the conditions we tested: the activity of Jak2^{Y637F} was decreased in the absence or presence of receptor and/or ligand activation and in the context of a Jak2 molecule that fails to bind most cytokine receptors. This is consistent with its position in the JH2 domain of Jak2, which constitutes a major regulator of Jak2 kinase activity (11, 24, 32, 37, 38). The phosphorylation of Tyr₆₃₇ during cytokine stimulation may thus act to alleviate JH2-mediated inhibition to promote the appropriate robust activation of Jak2 kinase activity. Indeed, while the phosphorylation of Tyr₆₃₇ requires the prior phosphorylation of Tyr_{1007/1008} (which promotes kinase activity by altering the conformation of the kinase domain), the phosphorylation and dephosphorylation of Tyr₆₃₇ progresses with a time course very similar to that of Tyr_{1007/1008}. That mutation of Tyr₆₃₇ partially suppresses the activating V617F JH2 mutation is also consistent with a role for Tyr₆₃₇ in reducing JH2-mediated inhibition of kinase function.

The activated phenotype of Jak2 molecules mutated at Ser₅₂₃, Tyr₃₁₇, and Tyr₅₇₀ suggests that the phosphorylation of each residue acts to inhibit the activity of Jak2. Ser₅₂₃ exhibits several differences compared to Tyr₃₁₇ and Tyr₅₇₀, however. Not only is the Ser₅₂₃-mediated inhibition weaker than that mediated by these other two sites (as shown by the relatively modest activation exhibited by its cognate Jak2 mutant), but Ser₅₂₃ is also highly (stoichiometrically) phosphorylated in the baseline state and not further phosphorylated during cytokine stimulation, at least in rapidly proliferating cells such as HEK293 and 32D cells (21). In contrast, Tyr₃₁₇ and Tyr₅₇₀ are phosphorylated upon cytokine receptor/Jak2 activation and more strongly inhibit Jak2 kinase activity. Interestingly, the phosphorylation of Tyr₃₁₇ is delayed compared to that of activating sites such as Tyr₆₃₇ and Tyr_{1007/1008}, depends upon prior phosphorylation of Tyr_{1007/1008} and an active conformation of Jak2, and remains highly phosphorylated even after these activating sites become relatively dephosphorylated.

Together, these observations suggest a model of phosphorylation-dependent regulation of Jak2 (Fig. 9) in which the

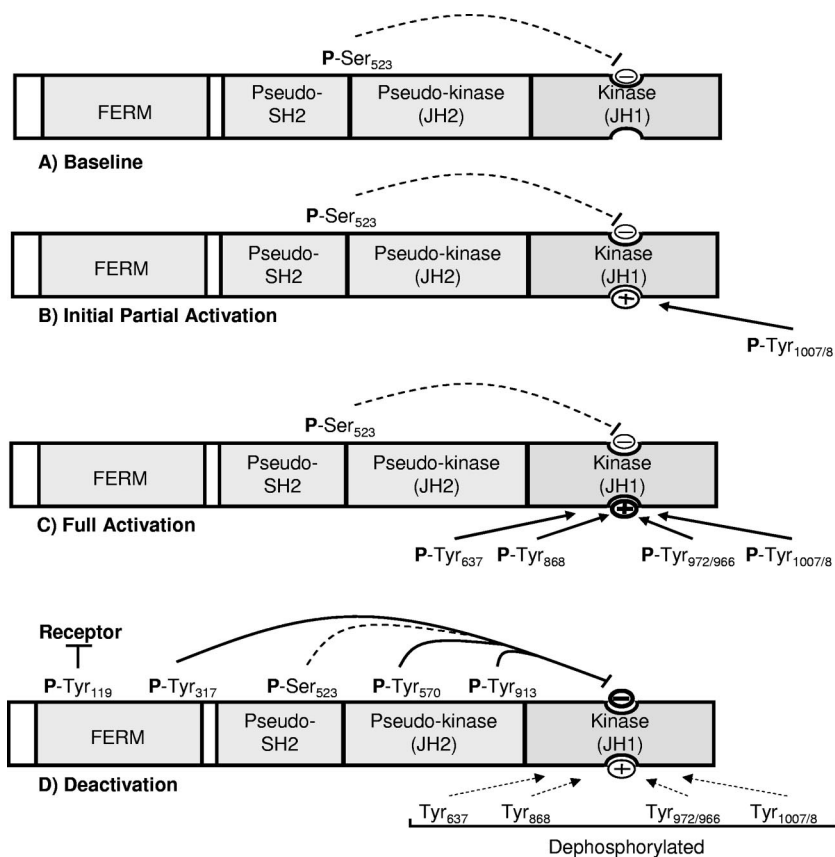


FIG. 9. Model of phosphorylation-dependent regulation of Jak2 activity. (A) Without stimulation, Jak2 is essentially unphosphorylated on most sites, although Tyr₅₂₃ is constitutively phosphorylated, providing a baseline of negative feedback to block kinase activity in the absence of stimulation. (B) Upon cytokine stimulation, Tyr_{1007/8} (the activation loop sites) are rapidly phosphorylated to promote partial activation of the kinase domain. (C) The phosphorylation of Tyr_{1007/1008} permits the phosphorylation of additional activating sites, including Tyr₆₃₇, as well as other sites in kinase domain, leading to full activation of Jak2. (D) After acute activation, strongly inhibitory sites, including Tyr₃₁₇, Tyr₅₇₀, Tyr₉₁₃, and Tyr₁₁₉, are phosphorylated to limit the extent of Jak2 activity, concomitant with the dephosphorylation of activating sites.

baseline phosphorylation of Ser₅₂₃ may serve to preserve the relative inactivity of Jak2 in the absence of cytokine stimulation. Since few forces promote the activation of Jak2 in the absence of stimulation, the relatively modest Ser₅₂₃-mediated inhibition suffices to suppress Jak2 activation in the absence of ligand binding, while the strong cytokine-mediated stimulus can overcome the weak inhibition by Ser₅₂₃. Upon cytokine stimulation, the rapid phosphorylation of Tyr_{1007/1008} alters conformation of the JH1 domain activation loop and partially activates the kinase, permitting the phosphorylation of Tyr₆₃₇, which promotes further Jak2 activation. Other sites in the kinase domain, such as Tyr₈₆₈, Tyr₉₆₆, and Tyr₉₇₂, may also be involved in fully activating Jak2 (27; Carter-Su and Argetsinger, unpublished).

The subsequent dephosphorylation of these stimulatory sites coincides with the relatively slower phosphorylation of strong inhibitory sites, such as Tyr₃₁₇ (and Tyr₅₇₀) to mediate feedback inhibition to limit the duration of the signal. Prolonged phosphorylation of the strong inhibitory sites also protects against continued high activity. The phosphorylation of Tyr₁₁₉ (which promotes receptor/Jak2 dissociation) and the inhibitory Tyr₉₁₃ in the kinase domain is also slightly delayed and more sustained compared to Tyr_{1007/1008} (13, 15), and the phosphor-

ylation of Tyr₁₁₉ and Tyr₉₁₃ likely functions together with Tyr₃₁₇ and Tyr₅₇₀ to limit the amplitude and duration of Jak2-dependent cytokine signaling.

Thus, a substantial subset of phosphorylation sites on Jak2 function to regulate Jak2 kinase activity. Each of these phosphorylation sites plays a specific role in the choreography of Jak2 activation and deactivation and, together, they ensure appropriate levels of Jak2 activity under a variety of circumstances. The functions of many other phosphorylation sites on Jak2 remain unknown. Although certain of these may possess relatively minor functions, others presumably contribute substantially to the regulation of Jak2 kinase activity and/or mediate as-yet-undiscovered downstream signals.

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