

Human *c-ros-1* Gene Homologous to the *v-ros* Sequence of UR2 Sarcoma Virus Encodes for a Transmembrane Receptorlike Molecule

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We isolated a human gene (designated *c-ros-1*) homologous to the *v-ros* sequence of UR2 sarcoma virus. Ten exons, 1,414 base pairs spanning 26 kilobases, contained a tyrosine kinase domain, a transmembrane domain, and a part of an extracellular domain carrying an N glycosylation site which was not acquired by UR2 sarcoma virus. The predicted structure of *c-ros-1* is unique among the *src* family and clearly distinct from the human insulin receptor.

UR2 sarcoma virus, a recent isolate of acutely transforming retrovirus of chickens (1), encodes for a fusion protein, p68^{gag-ros}, which has tyrosine-specific protein kinase activity (7). Nucleotide sequence analysis of the UR2 genome has revealed that the oncogene *v-ros* of UR2 carries a kinase domain homologous to those present in the oncogenes of the *src* family (12). The *v-ros* gene is considered to be derived from a cellular counterpart, proto-oncogene *c-ros* of chickens (15). Since the predicted chicken *c-ros* gene product, as well as the *v-ros* product, has a hydrophobic short stretch upstream of its kinase domain, it seems likely that the *c-ros* gene encodes for a transmembrane protein similar to the cell surface receptor for cell growth or differentiation factors (11, 12). Furthermore, recent reports have indicated that the deduced amino acid sequence of the kinase domain in the human insulin receptor (HIR) gene is highly homologous to the kinase domain in the *v-ros* sequence (6, 17). However, the phylogenetic conservation of the *c-ros* gene in mammalian species, including humans, and the relationship between the *c-ros* gene and the HIR gene have not yet been examined. In this study, we made an attempt to clarify these points.

High-molecular-weight genomic DNA was extracted from human placenta, mouse thymus, fish testis, *Drosophila melanogaster*, and yeast cells (*Saccharomyces cerevisiae*) and hybridized with a *v-ros*-specific probe (*Eco*RI-*Pvu*II 0.8-kilobase (kb) fragment; probe IV [see Fig. 2]). Under hybridization conditions of very relaxed stringency (20% formamide, 10% dextran sulfate, 1M NaCl; 37°C), we were able to detect clear bands in all DNAs examined, except for yeast DNA (Fig. 1). The *c-ros* sequence appeared to be conserved in vertebrate species from fish to mammals, including humans.

In human placenta DNA digested with the restriction endonuclease *Bam*HI, two discrete bands were detected at 15 and 10 kb (Fig. 1). Because these bands were observed only under very relaxed hybridization conditions, we molecularly cloned these *Bam*HI fragments, which may contain a portion(s) of the *c-ros* gene, before screening a large number of phages in a human genomic DNA library. With Charon 30

as a vector, four independent clones (two clones each from the 15- and 10-kb *Bam*HI fragments) were isolated by the method described by Benton and Davis (3) (Fig. 2). Partial DNA sequencing analysis by the dideoxy method (13) revealed that recombinant phage HYuros8 contained DNA sequences highly homologous to the *v-ros* sequence, whereas phage HYuros4 contained human sequences partially related to *v-ros* (data not shown). Both HYuros5 and HYuros1 were found to contain human sequences incidentally homologous to *v-ros*. Therefore, the human genes

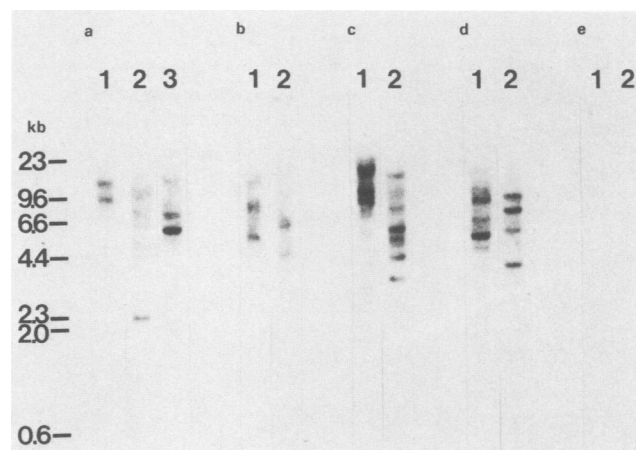


FIG. 1. Hybridization of a *v-ros*-specific probe to genomic DNAs. Cellular DNAs (10 μ g in panels a to c, 3 μ g in panel d, and 2 μ g in panel e) were digested with various endonucleases, electrophoresed on 0.8% agarose gels, and transferred to nitrocellulose filters (16). These filters were hybridized with a *v-ros*-specific probe (probe IV in Fig. 2) under hybridization conditions of low stringency (see text). Panels a to c were exposed to X-ray film at -70°C for 3 days; panels d and e were exposed for 6 days with an intensifying screen. Molecular weight markers were lambda DNAs digested with *Hind*III. Genomic DNAs used in Southern blot analyses were from human placenta (a), mouse thymus (b), fish testis (c), *D. melanogaster* (d), and *S. cerevisiae* (e) cells. These DNAs were digested with the following endonucleases. (a) Lanes: 1, *Bam*HI; 2, *Eco*RI; 3, *Hind*III. (b) Lanes: 1, *Bam*HI; 2, *Eco*RI. (c) Lanes: 1, *Bam*HI; 2, *Hind*III. (d) Lanes: 1, *Eco*RI; 2, *Hind*III. (e) Lanes: 1, *Bam*HI; 2, *Eco*RI.

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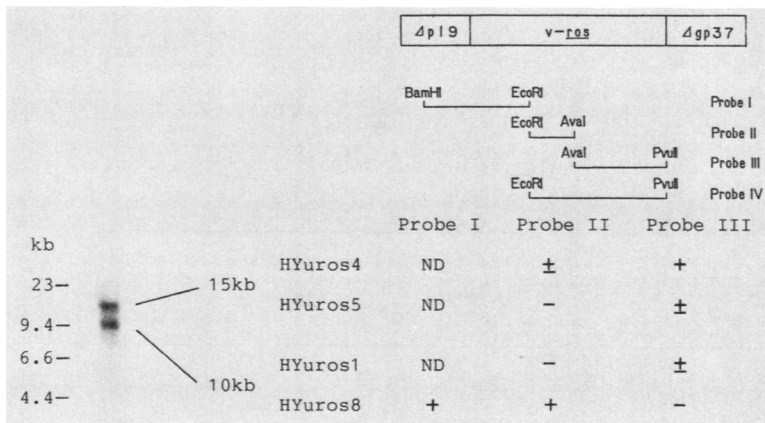


FIG. 2. Molecular cloning of human *c-ros* genes. Four independent clones were isolated. HYuros4 and HYuros5 were derived from phages recombined with 15-kb *Bam*HI fragments; the other two clones were derived from 10-kb *Bam*HI fragments. These four clones were hybridized with various *v-ros* probes under hybridization conditions of low stringency. Symbols: +, hybridization; ±, weak hybridization; -, no detected hybridization. ND, Not determined. Probes used are illustrated at the top of the figure. Molecular size markers (in kilobases) are indicated to the left of the gel.

found in HYuros8 and HYuros4 were designated human *c-ros-1* and human *c-ros-2*, respectively, and the human *c-ros-1* gene was further characterized in this study. The details of the analysis of the human *c-ros-2* gene will be described elsewhere.

We constructed a human genomic DNA library by the method described by Maniatis et al. (9), and gene walking of overlapping *c-ros-1* DNA sequences of about 60 kb was carried out. By restriction endonuclease mapping and by DNA sequencing of DNA fragments which hybridized with various *v-ros* probes, seven exons (exons 4 to 10) were found to encode for the entire kinase domain of this gene (Fig. 3a and 4). However, an approximately 240-base-pair region at the 5' end of the *v-ros* gene, including a possible transmembrane domain, was not detected by cross-hybridization between the human 60-kb DNA sequence and the *v-ros* sequence. We expected that the chicken *c-ros*

DNA fragment might be useful for isolating the transmembrane domain of the human *c-ros-1* gene more efficiently than could be done with the *v-ros* sequence as a probe. The ³²P-labeled 5' region of chicken *c-ros* DNA (5.2-kb *Eco*RI fragment [11]) was hybridized with the human DNA fragment upstream of the kinase domain (exon 1 in Fig. 3a), and then the nucleotide sequence of this region was determined. Although we did not obtain an exon(s) for a transmembrane domain in this region, to our surprise we found a new exon surrounded by a consensus splice acceptor site and a donor site (Fig. 3b); the predicted amino acid sequence had a potential site of N-linked glycosylation (Asn-Gly-Ser, amino acid residues 19 to 21; Fig. 4). In chicken *c-ros* DNA, an exon highly homologous to this human sequence at the level of predicted amino acids was also observed (Fig. 5). Since the amino acid sequence in this new exon (amino acid residues 1 to 63) was not present in the

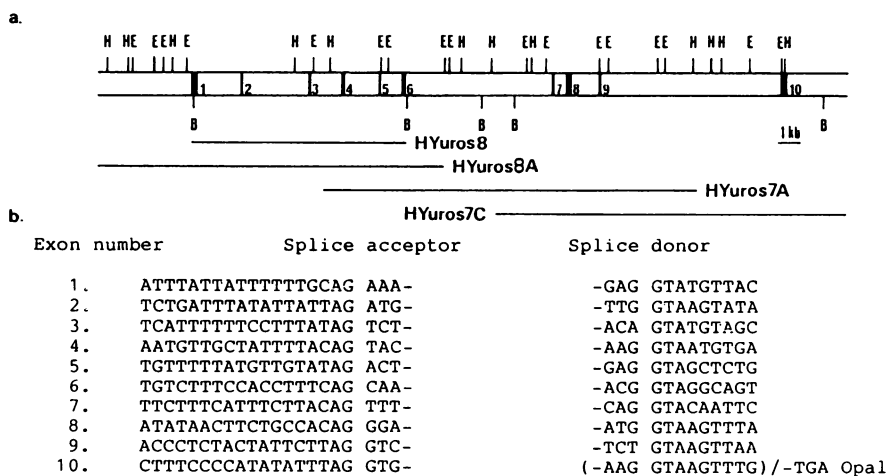


FIG. 3. Restriction map and gene organization of the human *c-ros-1* gene. (a) Four overlapping clones spanning 32-kb of cellular DNA are indicated. HYuros8 was a recombinant phage with Charon 30 *Bam*HI arms; the other clones, HYuros8A, HYuros7A, and HYuros7C, had Charon 4A *Eco*RI arms. The restriction map was determined by double or triple digestion with the following various endonucleases: B, *Bam*HI; E, *Eco*RI; and H, *Hind*III. The positions of exons in the human *c-ros-1* gene were determined by DNA sequencing. Black and white boxes indicate exons and introns, respectively. The numbers to the right of the black boxes indicate exon numbers. (b) Possible splice junctions in the human *c-ros-1* gene.

exon 1	Lys	Ser	Thr	Ser	Asn	Asn	Leu	Gln	Asn	10	Gln	Asn	Leu	Arg	Trp	Lys	Met	Thr	Phe	Asn	*	20	Gly	Ser	Cys	Ser	Ser	73
	A	AAG	AGC	ACT	TCA	AAT	AAT	TTA	CAG	AAC	CAG	AAT	TTA	AGG	TGG	AAG	ATG	ACA	TTT	AAT	GGG	TCC	TGC	AGT	AGT	148		
	Val	Cys	Thr	Trp	Lys	Ser	Lys	Asn	Leu	Lys	Gly	Ile	Phe	Gln	Phe	Arg	Val	Val	Ala	Ala	Asn	Asn	Leu	Gly	Phe	180		
	GTT	TGC	ACA	TGG	AAG	TCC	AAA	AAC	CTG	AAA	GGA	ATA	TTT	CAG	TTC	AGA	GTA	GTA	GCT	GCA	AAT	AAT	CTA	GGG	TTT	223		
50	Gly	Glu	Tyr	Ser	Gly	Ile	Ser	Glu	Asn	Ile	Ile	Leu	Val	Gly	Asp	Asp	Phe	Trp	Ile	Pro	Glu	Thr	Ser	Phe	Ile	298		
	GGT	GAA	TAT	AGT	GGA	ATC	AGT	GAG	AAT	ATT	ATA	TTA	GTT	GGA	GAT	GAT	TTT	TGG	ATA	CCA	GAA	ACA	AGT	TTC	ATA	375		
	Leu	Thr	Ile	Ile	Val	Gly	Ile	Phe	Leu	Val	Val	Thr	Ile	Pro	Leu	Thr	Phe	Val	Trp	His	Arg	Arg	Leu	Lys	Asn	448		
	CTT	ACT	ATT	ATA	GTT	GGA	ATA	TTT	CTG	GTT	GTT	ACA	ATC	CCA	CTG	ACC	TTT	GTC	TGG	CAT	AGA	AGA	TTA	AAG	AAI	523		
100	Gln	Lys	Ser	Ala	Lys	Glu	Gly	Val	Thr	Val	Leu	Ile	Asn	Glu	Asp	Lys	Glu	Leu	Ala	Glu	Leu	Arg	Gly	Leu	Ala	598		
	CAA	AAA	AGT	GCC	GAA	GGG	GTG	ACA	GTG	CTT	ATA	AAC	GAA	GAC	AAA	GAG	TTG	GCT	GAG	CTG	CGA	GGT	CTG	GCA	673			
	Ala	Gly	Val	Gly	Leu	Ala	Asn	Ala	Cys	Tyr	Gly	Ala	Ile	His	Thr	Leu	Pro	Thr	Gln	Glu	Glu	Ile	Glu	Asn	Leu	Pro	748	
	GCC	GGA	GTA	GGC	CTG	GCT	AAT	GCC	TGC	TAT	GCA	ATA	CAT	ACT	CTT	CCA	ACC	CAA	GAG	GAG	ATT	GAA	AAT	CTT	CCT	823		
150	Ala	Phe	Pro	Arg	Glu	Lys	Leu	Thr	Leu	Arg	Leu	Leu	Leu	Gly	Ser	Gly	Ala	Phe	Gly	Glu	Val	Tyr	Glu	Gly	Thr	898		
	GCC	TTC	CCT	CGG	GAA	AAA	CTG	ACT	CTG	CGT	CTC	TTG	CTG	GGA	AGT	GGA	GCC	TTT	GGA	GAA	GTG	TAT	GAA	GGA	ACA	973		
200	Ala	Val	Asp	Ile	Leu	Gly	Val	Gly	Ser	Gly	Glu	Ile	Lys	Val	Ala	Val	Lys	Thr	Leu	Lys	Lys	Gly	Ser	Thr	Asp	1048		
	GCA	GTG	GAC	ATC	TTA	GGA	GTT	GGA	AGT	GGA	GAA	ATC	AAA	GTA	GCA	GTG	AAG	ACT	TTG	AAG	AAG	GGT	TCC	ACA	GAC	1123		
250	Gln	Glu	Lys	Ile	Glu	Phe	Leu	Lys	Glu	Ala	His	Leu	Met	Ser	Lys	Phe	Asn	His	Pro	Asn	Ile	Leu	Lys	Gln	Leu	1198		
	CAG	GAG	AAG	ATT	GAA	TTC	CTG	AAG	GAG	GCA	CAT	CTG	ATG	AGC	AAA	TTT	AAT	CAT	CCC	AAC	ATT	CTG	AAG	CAG	CTT	1273		
300	Gly	Val	Cys	Leu	Asn	Glu	Pro	Gln	Tyr	Ile	Ile	Leu	Glu	Leu	Met	Glu	Gly	Gly	Asp	Leu	Met	Asp	Gly	Ile	Phe	1348		
	GGG	GTT	TGT	CTG	CTG	AAT	GAA	CCC	CAA	TAC	ATT	ATC	CTG	GAA	CTG	ATG	GAG	GGG	GGA	GAC	CTT	CTT	ACT	TAT	TTG	1417		
350	Arg	Lys	Ala	Arg	Met	Ala	Thr	Phe	Tyr	Gly	Pro	Leu	Leu	Thr	Leu	Val	Asp	Leu	Val	Asp	Leu	Cys	Val	Asp	Ile			
	CGT	AAA	GCC	CGG	ATG	GCA	ACG	TTT	TAT	GGT	CCT	TTA	CTC	ACC	TTG	GTT	GAC	CTT	GTA	GAC	CTG	TGT	GTA	GAT	ATT			
400	Ser	Lys	Gly	Cys	Val	Tyr	Leu	Glu	Arg	Met	His	Phe	Ile	His	Arg	Asp	Leu	Ala	Ala	Arg	Asn	Cys	Leu	Val	Ser			
	TCA	AAA	GGC	TGT	GTC	TAC	TTG	GAA	CGG	ATG	CAT	TTC	ATT	CAC	AGG	GAT	CTG	GCA	GCT	AGA	AAT	TGC	CTT	GTT	TCC			
450	Val	Lys	Asp	Tyr	Thr	Ser	Pro	Arg	Ile	Val	Lys	Ile	Gly	Asp	Phe	Gly	Leu	Ala	Arg	Asp	Ile	Tyr	Lys	Asn	Asp			
	GTG	AAA	GAC	TAT	ACC	AGT	CCA	CGG	ATA	GTG	AAG	ATT	GGA	GAC	TTT	GGA	CTC	GCC	AGA	GAC	ATC	TAT	AAA	AAT	GAT			
500	Tyr	Tyr	Arg	Lys	Arg	Gly	Glu	Gly	Leu	Leu	Pro	Val	Arg	Trp	Met	Ala	Pro	Glu	Ser	Leu	Met	Asp	Gly	Ile	Phe			
	TAC	TAT	AGA	AAG	AGA	GGG	GAA	GGC	CTG	CTC	CCA	GTT	CGG	TGG	ATG	GCT	CCA	GAA	AGT	TTG	ATG	GAT	GGA	ATC	TTC			
550	Thr	Thr	Gln	Ser	Asp	Val	Trp	Ser	Phe	Gly	Ile	Leu	Ile	Trp	Glu	Ile	Leu	Thr	Leu	Gly	His	Gln	Pro	Tyr	Pro			
	ACT	ACT	CAA	TCT	GAT	GTA	TGG	TCT	TTT	GGA	ATT	CTG	ATT	TGG	GAG	ATT	TTA	ACT	CTT	GGT	CAT	CAG	CCT	TAT	CCA			
600	Ala	His	Ser	Asn	Leu	Asp	Val	Leu	Asn	Tyr	Val	Gln	Thr	Gly	Gly	Arg	Leu	Glu	Pro	Pro	Arg	Asn	Cys	Pro	Asp			
	GCT	CAT	TCC	AAC	CTT	GAT	GTG	TTA	AAC	TAT	GTG	CAA	ACA	GGA	GGG	AGA	CTG	GAG	CCA	CCA	AGA	AAT	TGT	CCT	GAT			
650	Asp	Leu	Trp	Asn	Leu	Met	Thr	Gln	Cys	Trp	Ala	Gln	Glu	Pro	Asp	Gln	Arg	Pro	Thr	Phe	His	Arg	Ile	Gln	Asp			
	GAT	CTG	TGG	AAT	TTA	ATG	ACC	CAG	TGC	TGG	GCT	CAA	GAA	CCC	GAC	CAA	AGA	CCT	ACT	TTT	CAT	AGA	ATT	CAG	GAC			
700	Gln	Leu	Gln	Leu	Phe	Arg	Asn	Phe	Phe	Leu	Asn	Ser	Ile	Tyr	Lys	Ser	Arg	Asp	Glu	Ala	Asn	Asn	Ser	Gly	Val			
	CAA	CTT	CAG	TTC	TTC	AGA	AAT	TTT	TTC	TTA	AAT	AGC	ATT	TAT	AAG	TCC	AGA	GAT	GAA	GCA	AAC	AAC	AGT	GGA	GTC			
750	Ile	Asn	Glu	Ser	Phe	Glu	Gly	Lys	Phe	Asp	Ser	Ser	Glu	Phe	Ser	Ser	Phe	Arg	Cys	Thr	Val	Asn	Opal					
	ATA	AAT	GAA	AGC	TTT	GAA	GCT	AAG	TTT	GAT	TCT	TCA	GAA	TTT	TCT	AGT	TTT	CGC	TGC	ACT	GTC	AAC	TGA					

FIG. 4. Nucleotide sequence and predicted amino acid sequence of the human *c-ros-1* gene. The putative transmembrane domain and kinase domain are underlined and boxed, respectively. A potential site of N-linked glycosylation is indicated by an asterisk. The horizontal arrows above the amino acid sequences indicate the junctions between exons and introns. The nucleotides above the dashed line indicate a possible splice donor site. Ala, Alanine; Cys, cysteine; Asp, aspartic acid; Glu, glutamic acid; Phe, phenylalanine; Gly, glycine; His, histidine; Ile, isoleucine; Lys, lysine; Leu, leucine; Met, methionine; Asn, asparagine; Pro, proline; Gln, glutamine; Arg, arginine; Ser, serine; Thr, threonine; Val, valine; Trp, tryptophan; Tyr, tyrosine.

v-ros sequence of UR2 sarcoma virus, it seemed most likely that this exon belonged to the extracellular domain of the *c-ros* gene not acquired in the viral genome.

By nucleotide sequencing of the entire cellular DNA of about 6 kb in length from exon 1 to exon 4 in Fig. 3a, two exons, exon 2 and exon 3, were detected. The predicted amino acid sequence of exon 2 showed an extremely hydrophobic stretch of 21 amino acids; that of exon 3 carried a sequence 63% homologous to the corresponding exon of the chicken *c-ros* gene (Fig. 4, Table 1). Although the nucleotide sequence of exon 2 greatly diverged from that of the chicken *c-ros* transmembrane domain and the peptide length was 2 amino acids shorter than that in chickens, we consider this hydrophobic stretch to be the transmembrane domain of the human *c-ros-1* gene because of its partial homology to the hydrophobic amino acid sequence of the chicken *c-ros* gene and because its length is sufficient to pass through the lipid bilayer of the cell membrane. The extents of nucleotide

homology of the regions of exon 2 and exon 3 with the corresponding regions of the chicken *c-ros* gene were 52 and 60%, respectively. This weak homology may explain the failure to detect cross-hybridization between these sequences and the *v-ros* sequence by Southern blot analysis. Such a high degree of heterogeneity in the nucleotide sequence outside the kinase domain of an oncogene between avian and human species has also been reported in the case of the *c-src* gene; exon 3 in the human *c-src* gene could not be detected by cross-hybridization using the *v-src* sequence as a probe (8).

Ten exons of the human *c-ros-1* gene were identified within this 26-kb human DNA separated by 1- to 6-kb-long introns (Fig. 3a). The entire coding sequence and the predicted amino acid sequence of the human *c-ros-1* gene are shown in Fig. 4. The kinase domain of the human *c-ros-1* gene was found in the sequences from exon 4 to a part of exon 10 (Fig. 4). In these exons, the structure was highly



FIG. 5. Comparison of the amino acid sequences among the human *c-ros-1* gene, the chicken *c-ros* gene, and the HIR gene. Symbols: colon (:), identical amino acid; +, conserved amino acid; -, deletion of amino acid.

homologous to chicken *c-ros* and *v-ros* genes not only in the DNA sequence but also in the predicted amino acid sequence, except for exon 10 (Table 1, Fig. 5). Furthermore, the predicted products of the human *c-ros-1* gene and the chicken *c-ros* gene shared similar inserts of 2 to 5 amino acids (amino acid residues 252 to 254, 260 to 261, and 299 to 303) which were not present in any other members of the *src* family, and the splice junctions in these two genes were completely matched to each other (11). From these results, we conclude that the human *c-ros-1* gene is a cellular DNA homolog of *v-ros* in the human genome. Recent studies on the structure of the HIR gene have shown that the kinase domain of the HIR gene deduced from the cDNA sequence is more homologous to that of *v-ros* than it is to those of other *src* family members. However, by comparison of the

predicted amino acids of the human *c-ros-1* gene and the HIR gene, these two molecules were demonstrated to be clearly different from each other; homology in the level of amino acids in the kinase domain was 48.5% (Fig. 5).

In the overall structure, the human *c-ros-1* gene carried an extracellular domain with a potential site of N-linked glycosylation, a hydrophobic 24-amino acid stretch, and a tyrosine kinase domain. These structural organizations are similar to those of the *c-erbB* (the gene for epidermal growth factor receptor), the *c-fms* (the gene for macrophage colony-stimulating-factor receptor), and the HIR genes (5, 6, 14, 17, 18). These results strongly suggest that the human *c-ros-1* gene encodes for a transmembrane molecule which may function as a receptor for a cell growth or differentiation factor(s). Recently isolated transforming genes, *neulerbB2* and *oncD*, appear to be derived from the same category of receptor-type proto-oncogene (2, 4, 10, 19).

The biological function of the *c-ros* gene product and the significance of the *c-ros* gene in tumorigenicity in animals remain to be elucidated. Expression of the *c-ros* gene in 7- to 14-day-old healthy chickens was strongly repressed in many tissues, but two to three copies per cell of *c-ros* RNA were detected in kidneys by liquid hybridization and Northern blotting methods (11, 15). These results might indicate that the *c-ros* gene has a function in a limited stage of development or in a particular cell population in some tissue such as the kidney. Molecularly cloned human *c-ros-1* DNA may be very useful for examining the expression or abnormalities of this gene in normal or malignant tissues.

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TABLE 1. Homology between the human *c-ros-1* gene and the chicken *c-ros* gene in terms of nucleotide sequence and predicted amino acid sequence

Human <i>c-ros-1</i> exon no.	No. of nucleotides identical to chicken <i>c-ros</i> (n)	% Homology	No. of amino acids identical to chicken <i>c-ros</i> (n)	% Homology
1	121 ^a (191) ^b	63.4 ^a	32 ^a (63) ^b	50.8
2	44 (84)	52.4	9 (28)	32.1
3	81 (136)	59.6	29 (46)	63.0
4	121 (163)	74.2	42 (54)	77.8
5	55 (65)	84.6	19 (22)	86.4
6	102 (130)	78.5	34 (43)	79.1
7	68 (98)	69.4	24 (33)	72.7
8	156 (201)	77.6	57 (67)	85.1
9	96 (135)	71.1	29 (45)	64.4
10	100 (211)	47.4	25 (70)	35.7

^a This possible extracellular domain in the chicken *c-ros* gene is not present in the *c-ros* sequence reported by Neckameyer et al. (11) and has been sequenced in this study.

^b n, Total number of nucleotides or amino acids in each exon.

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